

## CLINICAL PRACTICE

**The assessment of erythrocyte deformability by filtration rate**

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*Institute of Physiology, Faculty of Medicine, Comenius University, Bratislava, Slovakia. mataseje@fmed.uniba.sk***Abstract**

The aim of the study was to monitor the erythrocyte deformability as one of important factors securing the appropriate tissues perfusion in healthy subjects and diabetic patients with insulin dependent diabetes mellitus. Erythrocyte deformability was determined by the method of filtration and centrifugation, and the erythrocyte filtrability was calculated as a percentage of filtered erythrocytes out of the number of erythrocytes counted before centrifugation. Diluted blood suspensions were filtered by centrifugation through membrane filters with pores of 5  $\mu\text{m}$  in diameter. The speed and duration of centrifugation of 1400 rpm and 5 min respectively were selected as the best ones for filtration due to the lowest value of the coefficient of variance.

The values of the arithmetic mean and standard deviation of erythrocyte filtrability were  $72.2 \pm 7.9$  % in normal subjects. In the group of diabetic patients with the long history of insulin therapy the values amounted to  $69.1 \pm 4.4$  %. In diabetic patients, the average value of blood glucose was  $11.7 \text{ mmol.L}^{-1}$ , and the glycated haemoglobin concentration reached 9.04 %. From the viewpoint of reference values, these facts indicate good compensation of diabetes mellitus. Other haematological values, namely erythrocyte count, haematocrit value, haemoglobin concentration and mean cell volume were within normal reference ranges. No difference between the groups of healthy subjects and diabetic patients was found. (Fig. 4, Ref. 18.)

**Key words:** erythrocyte deformability, erythrocyte filtration, diabetes mellitus.

Erythrocyte deformability (ED) is one of important physiological factors securing the appropriate tissues perfusion. To realize this function, erythrocytes have to deform their shapes by reducing their stationary diameter of 7–8  $\mu\text{m}$  to diameters as small as 1.5  $\mu\text{m}$  in order to be able to traverse the narrow capillary beds and splenic sinusoids. The deformability of normal human erythrocytes is the consequence of their low cytoplasmic viscosity, their excess of membrane surface in relation to cell volume and their viscoelastic properties of cell membrane. The erythrocyte flexibility is determined by the combination of these factors. Loss of deformability may arise from impairment of any of these properties or their combinations (1).

Erythrocyte deformability changes play an important role in the pathophysiology of some diseases, e.g. haematologic diseases (2, 3), diabetes mellitus (4, 5, 6), ischaemic heart disease (7, 8), stroke and hypertension (9), as well as in coincidence with hard exercise (10, 11). Because of the diagnostic importance of ED determination, several methods have been developed enabling to measure it.

There are various methods and instruments available for ED measuring: methods, in which erythrocyte suspensions are sheared

in geometric systems (2, 6), e.g. viscometry and laser diffraction ellipsometry (10, 12, 13) and methods in which erythrocytes are made to traverse narrow channels – micropipettes (14) or filter pores (7, 15). To study the erythrocyte microrheology, the method of cation-osmotic haemolysis has been developed (16) and used in many experimental and clinical studies (17, 18).

Erythrocyte filtration is a technique widely used in clinical assessment of ED. In this work the method of erythrocyte filtration was selected for its low costs and quickness of procedure.

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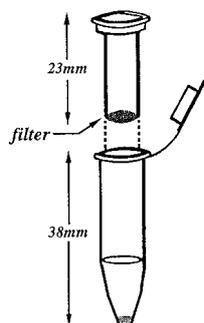


Fig. 1. An ultrafree membrane filter.

### Materials and methods

Two groups of students were examined. The first consisted of 7 healthy subjects, non-diabetics, aged 20 to 23 and the second one included 16 patients with insulin dependent diabetes mellitus (IDDM) aged 21 to 31. They had been treated by insulin for 14 years on the average.

The haematologic analysis and ED determination were made from venous blood samples taken into sterile syringes with potassium-EDTA as anticoagulant. Blood suspensions for erythrocyte filtration were prepared by diluting 20  $\mu\text{L}$  of anticoagulated blood with 10 mL of diluting solution RR I (Bachem, Slovakia), which contains boric acid and sodium chloride. The dilution of whole blood in suspension was 1:500. An aliquot of 100  $\mu\text{L}$  of each erythrocyte suspension was taken to 10 ml of diluting solution to determine the erythrocyte count (RBC count), haematocrit value, and mean cell volume by the semiautomated cell counter Picoscale 5 (Medicor, Hungary). Another 200  $\mu\text{L}$  aliquot of each suspension was pipetted into the centrifugal tube (Fig. 1) equipped with membrane filter with average pores of 5  $\mu\text{m}$  in diameter (Ultrafree MC, Millipore, Japan). The tubes with blood suspensions were centrifuged by microcentrifuge Micro 20 (Hettich,

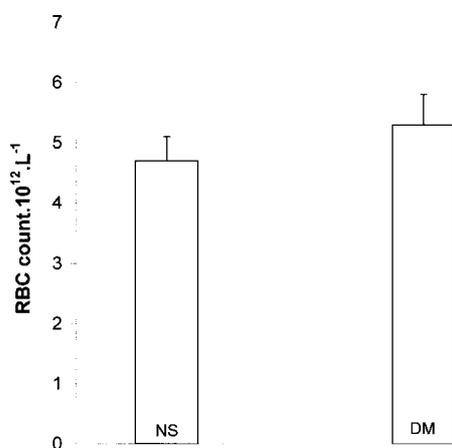


Fig. 2. Red blood cell count in normal subjects (NS) and patients with diabetes mellitus (DM).

Deutschland) at 1400 rpm for 5 minutes. After centrifugation, in the course of which the erythrocytes were passing through narrow pores of filter, each supernatant was well mixed to diffuse filtered erythrocytes uniformly. The filtrated suspensions were appropriately diluted and counted by the cell counter to determine the haematological values described above. The haemoglobin concentration was measured by the cyanmethaemoglobin method and reticulocytes were counted microscopically. All measurements were carried out at the laboratory temperature of 22–24 °C and performed no later than four hours after venepuncture.

ED assessment was calculated on the basis of filtration rate:

$$\text{filtration rate} = \frac{\text{RBC count after centrifugation}}{\text{RBC count before centrifugation}} \times 100 (\%)$$

Means and standard deviations of filtration measurements were used to calculate the coefficient of variation (CV):

$$\text{CV} = \frac{\text{standard deviation}}{\text{mean}} \times 100$$

To find the best conditions for filtration, the means and variation coefficients were studied under different centrifugation conditions with eighth aliquots. The tubes were centrifuged at 1200 rpm/5 min, 1200 rpm/10 min, 1300 rpm/5 min and 1400 rpm/5 min. The mean with the standard deviation of filtration rate and the coefficient of variation were calculated for each centrifugation condition. The lowest CV 7.1 was obtained under centrifugation of 1400 rpm/5 min. Due to this result, all experiments were performed under the same conditions.

The experimental data were statistically tested by non-parametric Kolmogorov–Smirnov test.

### Results

In the group of normal subjects, the average erythrocyte count was  $4.7 \pm 0.4 \cdot 10^{12} \cdot \text{L}^{-1}$  (Fig. 2). After the erythrocytes filtration, the filtration rate  $72.2 \pm 7.9 \%$  was calculated (Fig. 3). Other haematologic values were assessed: mean cell volume was  $82.7 \pm 6.4 \cdot 10^{-15} \text{L}$  (Fig. 4), haemoglobin concentration  $153.5 \pm 16 \text{g} \cdot \text{L}^{-1}$  and haematocrit value  $39.1 \pm 4.0 \%$  were in the range of physiological values. Reticulocytes count presents  $0.90 \pm 0.2 \%$  of the total erythrocyte count in normal subjects.

In the group of diabetic patients the average glucose value was  $11.7 \text{mmol} \cdot \text{L}^{-1}$  and glycated haemoglobin concentration was 9.04 %. This fact refers to good compensation of diabetes from the viewpoint of physiological values. In this group, the average value of erythrocyte count was higher  $5.3 \pm 0.5 \cdot 10^{12} \cdot \text{L}^{-1}$  ( $p=0.06$ ) against the control group (Fig. 2). The obtained erythrocyte filtration value of  $69.1 \pm 4.4 \%$  (Fig. 3) was lower by 4.3 % in comparison with the group of normal subjects. Other haematological variables that were assessed as average values – mean cell volume  $87.8 \pm 3.5 \cdot 10^{-15} \text{L}$  (Fig. 4), haemoglobin concentration  $172.0 \pm 13 \text{g} \cdot \text{L}^{-1}$  and haematocrit value

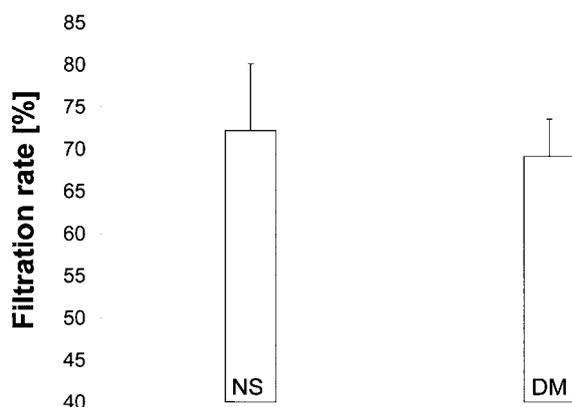


Fig. 3. Filtration rate in normal subjects (NS) and patients with diabetes mellitus (DM).

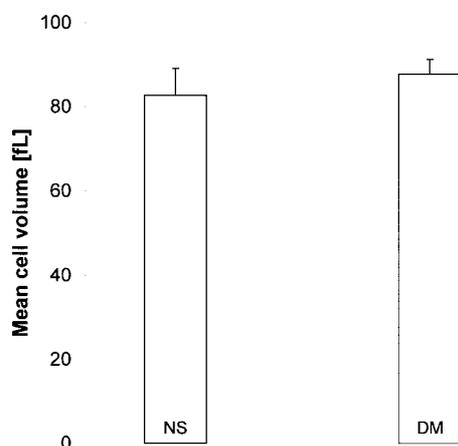


Fig. 4. Mean cell volume in normal subjects (NS) and patients with diabetes mellitus (DM).

46.7±3.1 %. Reticulocyte count from the total erythrocyte count being 0.90±0.2 % was the same as in the group of normal subjects.

## Discussion

The presented filtration method for ED determination was tested under different centrifugation conditions. The best condition found for centrifugation was 1400 rpm/5 min, as it yielded the lowest value of variation coefficient of 7.1. Membranal filters with pores of 5 µm in diameter were used according to the recommendations of International Committee for Standardization in Haematology, that proposes the pores to range between 3–5 µm in diameter (1).

Our results of erythrocyte filtration rate correspond with the literary data (4, 5). It was found out that significantly lower erythrocyte filtration rate in patients with IDDM was not significantly lower in comparison to the control group. This fact can be explained by the protective effect of long-term insulin therapy and good compensation of diabetes mellitus. Linderkamp et al (6) tested the influence of insulin therapy on ED. In children with IDDM before the initiation of insulin treatment, ED was decreased by 28,

after 5 to 8 years of insulin treatment ED improved and decreased by 17 when compared with healthy children. This outcome refers to the fact that long-term insulin therapy has a positive effect on ED. Our results also refer to the favourable effect of long-term insulin therapy. Erythrocyte deformability value is probably influenced by the assessed higher values of mean cell volume, haemoglobin concentration and haematocrit value in the group of diabetic patients.

The application of the filtration method for the assessment of ED will contribute to the specification of diagnostic and therapeutic procedures in clinical practice.

## References

1. Bull BS, Chien S, Dormandy JA, Kiesewetter H, Lewis SM. Guidelines for Measurement of Blood Viscosity and Erythrocyte Deformability. *Clin Hemorheol* 1986; 6: 439–453.
2. Dondorp AM, Chotivanich KT, Fucharoen S, Silamut K, Vreeken J, Kager PA, White NJ. Red cell deformability, splenic function and anaemia in thalassaemia. *Brit J Haematol* 1999; 105: 505–508.
3. Ramakrishnan S, Grebe R, Singh M, Schmid-Schonbein H. Evaluation of hemorheological risk factor profile in plasmacytoma patients. *Clin Hemorheol Microcirc* 1999; 20: 11–19.
4. Okada M, Matsuto T, Sugita O, Kimura S, Matsumoto H, Okada T. A simple method to measure red blood cell deformability by centrifugation. *La Presse Medicale* 1994; 23: 1613–1615.
5. Tsukada K, Sekizuka E, Oshio C, Minamitani H. Direct measurement of erythrocyte deformability in diabetes mellitus with a transparent microchannel capillary model and high-speed video camera system. *Microvasc Res* 2001; 61: 231–239.
6. Linderkamp O, Ruef P, Zilow EP, Hoffmann GF. Impaired deformability of erythrocytes and neutrophils in children with newly diagnosed insulin-dependent diabetes mellitus. *Diabetologia* 1999; 42: 865–869.
7. Penco M, Romano S, Dagianti A Jr, Tozzi-Ciancarelli MG, Dagianti A. Modifications of whole blood filterability during acute myocardial infarction. *Clin Hemorheol Microcirc* 2000; 22: 153–159.
8. Turchetti V, Leoncini F, De Matteis C, Trabalzini L, Guerrini M, Forconi S. Evaluation of erythrocyte morphology as deformability index in patients suffering from vascular diseases, with or without diabetes mellitus: correlation with blood viscosity and intraerythrocytic calcium. *Clin Hemorheol Microcirc* 1998; 18: 141–149.
9. Cicco G, Pirrelli A. Red blood cell (RBC) deformability, RBC aggregability and tissue oxygenation in hypertension. *Clin Hemorheol Microcirc* 1999 21: 169–177.
10. Smith JA, Martin DT, Telford RD, Ballas SK. Greater erythrocyte deformability in world-class endurance athletes. *Amer J Physiol* 1999; 276: H2188–2193.
11. Nakano T, Wada Z, Matsumura S. Membrane lipid components associated with increased filterability of erythrocytes from long-distance runners. *Clin Hemorheol Microcirc* 2001; 24: 85–92.
12. Wang X, Zhao H, Zhuang FY, Stolz JF. Measurement of erythrocyte deformability by two laser diffraction methods. *Clin Hemorheol Microcirc* 1999; 21: 291–295.
13. Yao W, Wen Z, Yan Z, Sun D, Ka W, Xie L, Chien S. Low viscosity ektacytometry and its validation tested by flow chamber. *J Biomech* 2001; 34: 1501–1509.
14. Ruef P, Linderkamp O. Deformability and geometry of neonatal erythrocytes with irregular shapes. *Pediatr Res* 1999; 45: 114–119.
15. Moriguchi T, Takasugi N, Itakura Z. The effects of aged garlic extract on lipid peroxidation and the deformability of erythrocytes. *J Nutr* 2001; 131: 1016S–1019S.
16. Nicák A, Mojžiš J. Differences in the haemolytic action of mercury ions on human and rat erythrocytes with relationship to the concentration of Na<sup>+</sup> and glucose in vitro. *Comp Haematol Int* 1992; 2: 84–86.
17. Pomfy M, Nicák A. The effect of global cerebral ischemia on erythrocyte deformability and cerebral microcirculation in dogs. *Bratisl Lek Listy* 1996; 97: 365–368.
18. Mojžiš J, Nicák A, Troščák M, Kozák I, Mirossay L. Cation — osmotic haemolysis in stroke patients. *Comp Haematol Int* 1999; 9: 83–85.

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