

REVIEW

Adenosine and cardioprotection: What can we learn from nature's genetic polymorphism?

Skalova K, Luptak I, Matuskova J, Turcani M, Hulin I

Institute of Pathophysiology, Faculty of Medicine, Comenius University, Bratislava, Slovakia. bl@fmed.uniba.sk

Abstract

Adenosine is an endogenous nucleoside that has been shown to be beneficial for the myocardium in different settings by a large number of experimental studies. In this article, we 1) outline adenosine's metabolic pathways, 2) address cardioprotective properties of adenosine, and 3) discuss possible implications of the two recently published clinical studies disclosing a positive effect of adenosine monophosphate deaminase 1 (AMPD1) gene mutation on cardiovascular survival in heart failure and ischemic heart disease. (Fig. 2, Ref. 84.)

Key words: adenosine, cardioprotection, AMPD1 gene mutation.

Adenosine is an endogenously occurring nucleoside that participates in modulation of numerous physiological functions. In relation to cardiovascular system, it was in 1929 when Drury and Szent-Györgyi first described its bradycardic, vasodilatory and blood pressure lowering effects (1). A large number of studies published over the last two decades, offering experimental evidence adenosine may be beneficial for the myocardium in different settings, has lead to the tenet that this nucleoside is a cardioprotective metabolite (2–8). However, the extremely short half-life of adenosine and tachyphylaxis after its prolonged administration dissuaded investigators from exploring potentially favorable effects of up-modulation of this substance on a long-term basis in patients with heart diseases. In this article, we will 1) outline adenosine's metabolic pathways, 2) briefly address cardioprotective properties of adenosine, and 3) discuss the possible revolutionary implications that may arise from the two recently published clinical studies disclosing a positive effect of adenosine monophosphate deaminase 1 (AMPD1) gene mutation on cardiovascular survival in heart failure (9) and ischemic heart disease (10).

Metabolism of adenosine

Adenosine is a purine nucleoside comprised of adenine and ribose joined by a glycosidic bound (Fig. 1). It is formed in tissues via dephosphorylation of adenosine 5'-monophosphate (AMP) by 5'-nucleotidase or via hydrolysis of S-adenosylhomocysteine (SAH) by SAH-hydrolase, the former being the major source of cardiac adenosine production.

Adenosine's precursor AMP is produced mainly by adenosine 5'-triphosphate (ATP) breakdown, although production from cyclic AMP (cAMP) by phosphodiesterase has also been documented in the cardiac fibroblasts (11) and vascular smooth muscle cells (12). AMP can be deaminated by AMP deaminase to form inosine monophosphate (IMP), or dephosphorylated by 5'-nucleotidase to form adenosine (5) (Fig. 2). Two different forms of 5'-nucleotidase play an important role in the adenosine formation (13): the AMP-selective cytosolic 5'-nucleotidase (cN-I) (14–16) and ecto-5'-nucleotidase (17). The ecto-5'-nucleotidase is expressed on the outer membrane of many cell types in the heart, including pericytes, fibroblasts (18), endothelial cells, smooth muscle cells (19) and cardiomyocytes (20, 21). In the heart, adenosine is thus formed both intracellular in cardiomyocytes (22), coronary smooth muscle cells (23) and endothelial cells (24), and extracellular from adenosine nucleotides released from cardiomyocytes, endothelial cells (25) and blood elements.

Institute of Pathophysiology, Faculty of Medicine, Comenius University, Bratislava, Slovakia, and Department of Physiology, Kuwait University, Kuwait

Address for correspondence: K. Skalova, MD, Institute of Pathophysiology, LFUK, Spitalska 24, SK-813 72 Bratislava 1, Slovakia.
Phone: +421.2.52965400 Fax: +1508.856.4571

Acknowledgement: The authors of this article were supported by the grant of Scientific Agency of Ministry of Education of Slovak Republic No. 1/7492/20.

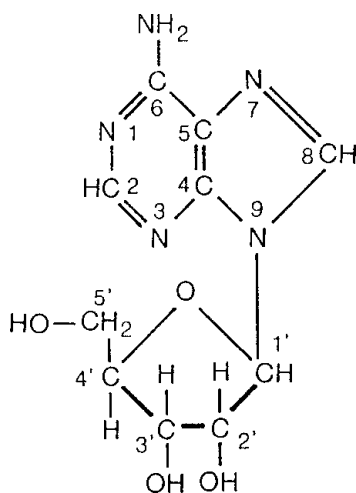


Fig. 1. Structure of adenosine.

Adenosine crosses the cell membranes by facilitated diffusion. It can be rapidly inactivated by phosphorylation to AMP by adenosine kinase, or deaminated to inosine by adenosine deaminase (26). The details about compartmentalization and regulation of adenosine formation in the respective compartments, as well as the transport of this nucleoside out of and into cells is a subject of ongoing experimental research (27, 28) and mathematical modeling (29, 30).

In tissues that are dependent on oxidative phosphorylation such as the heart, a decreased oxygen supply relative to the workload leads to an energy-deficient state. This situation shifts the metabolism of adenine nucleotides towards the degradation of ATP with augmented generation of adenosine 5'-diphosphate (ADP) and thence, AMP, the direct substrate for adenosine production. Moreover, cN-I and ecto-5'-nucleotidase have been noted to be activated by ischemia (31), the later also by α -1-adrenergic receptor stimulation (32), which would further contribute to the increased adenosine formation in the energy-deficient state. The total production of adenosine in the heart has been documented to be markedly elevated in hypoxia and ischemia (5), both in experiments (33–36), and in the clinical setting of ischemic heart disease (37) and heart failure (38).

In summary, the most important source of adenosine in cells and in the interstitial tissue is the “ATP→ADP→AMP→adenosine” pathway. Hypoxia and ischemia strongly enhances adenosine production. Thus, adenosine can be considered a signal of inadequate oxygen supply for the tissue (39, 40).

Cardioprotection by adenosine

The majority of adenosine's effects can be viewed as directed towards restoring the energy balance in the setting of relative shortage of oxygen supply. It acts to decrease the workload of the heart: negative dromo-, chrono- and inotropic effects, as well as to increase the oxygen supply for the tissues: dilatation of

coronary arteries, vasodilatation in other vascular beds, stimulation of breathing (2, 40). These properties of adenosine lead to the concept this nucleoside acts in the cardiovascular system as a retaliatory metabolic counteracting the mismatch between energy supply and demand and this way protecting the heart (41). Furthermore, it has been noted in the past decade the beneficial effects of adenosine surpasses its retaliatory function (3, 5, 7, 8). Adenosine has been shown to be one of the major triggers involved in the mechanism of ischemic preconditioning, the most potent mechanism protecting the heart against ischemia known to date.

Ischemic preconditioning is a striking phenomenon that has been first described in 1986 by Murry et al: a significant reduction in infarct size has been observed when the heart was previously subjected to shorter episodes of ischemia (42). Early and delayed phases of preconditioning have been described, where the initial ischemic episode conveys cardioprotection against a following ischemic event that can occur either immediately – phase of preconditioning!, or 24–72 hours later – “the delayed phase”, or “second window of preconditioning.” Although much work is still needed in order to elucidate the entire sequence of events responsible for the cardioprotective effect of short-term ischemia, the mechanism of ischemic preconditioning has been explained in part. Namely distinct extracellular triggers have been detected and intracellular mediators and downstream signaling pathways of preconditioning described (43).

Adenosine is quickly released into extracellular fluid during ischemia and has been shown to be one of the most important triggers implicated in ischemic preconditioning. Infusion of adenosine (44, 45), as well as adenosine A1 (46–49) and A3 (50–54) receptor agonists, mimics the preconditioning effect of short-term ischemia, and protects the heart both against early and delayed ischemic insult (55–59). Moreover, these effects can be blocked by adenosine receptor antagonists (60). The downstream signaling pathway activated by A1 and/or A3 adenosine receptors, both of them G-protein coupled acting through inhibition of adenylyl cyclase (A3 possibly also through increase in activity of phospholipase C), involve opening of mitochondrial K_{ATP} channels (61), generation of free radicals (62), translocation of the epsilon isoform of protein kinase C (63, 64) and activation of the family of mitogen activated protein kinases (65). The latter induce the transcription of certain enzymes, such as superoxide dismutase, inducible NO synthase (66) and heat-shock proteins (67, 68), which can account for the second window of protection in the delayed phase of preconditioning.

Clinical studies on AMPD1 gene mutation

On the experimental level, extensive evidence has been gathered that adenosine can exert cardioprotective effects. However, clinical studies analyzing its potential cardioprotective effects in humans have been limited to trials using short-term administration of adenosine in acute ischemia (69), preconditioning before percutaneous transluminal interventions (45, 70) and coronary artery bypass grafting (71), and its use in cardioplegic solutions

vival in AMPD1 gene mutation carriers documented in the two aforementioned studies. The decreased AMPD1 activity in the skeletal muscle of patients heterozygous or homozygous for AMPD1 gene mutation presumably leads to accumulation of AMP in their skeletal muscle with the shift of AMP metabolism towards adenosine formation by 5'-nucleotidase (Fig. 2). Given the previously mentioned cardioprotective properties of adenosine, the increased production of this nucleoside appears to be the most plausible explanation for the improved survival of cardiac patients with AMPD1 gene mutation (9, 10, 75). Loh with co-authors suggested that in AMPD1 deficiency circulating levels of adenosine are increased due to its augmented release from the skeletal muscle (9). Yet, the known short half-life of adenosine in human blood lead to doubts if it is the increased formation in skeletal muscle that accounts for the cardioprotective results, and rather an enhanced local production in the cardiac muscle has been proposed (75). However, no skeletal, myocardial or circulating adenosine levels were determined in either study. What the kinetics of adenosine in the setting of AMPD1 deficiency is, and how the nucleoside exerts its protective effects on the cardiac tissue, remains an intriguing question.

We propose the hypothesis that the critical characteristic of adenosine kinetics in AMPD1 deficiency that results in significant cardioprotection is the intermittent manner of the adenosine level increase. There are several lines of evidence to support this.

First, skeletal muscle adenosine levels at rest detected by muscle biopsy from patients with myoadenylate deaminase deficiency have been reported to be comparable to those of non-carriers of AMPD1 mutation (76). If the skeletal muscle is the major site of AMPD1 expression, and its deficiency does not lead to locally increased adenosine levels at this site during resting condition, there is no reason to expect adenosine levels in circulation or in the myocardium would be increased in a continuous fashion. On the other hand, exercise has been shown to elevate adenosine levels in skeletal muscle biopsies up to 16-fold in AMPD1 deficiency comparing to the 2-fold increase with exercise in AMPD1 mutation non-carriers (76), which is not surprising. The increased energy demand during physical activity leads to increased degradation of ATP in skeletal muscle and accumulation of large amounts of AMP that, in deficiency of AMPD1, can be metabolized only by 5'-nucleotidase to form adenosine. This purine is one of chemical mediators of pain, and this could partly explain the clinical observation that patients with myoadenylate deaminase deficiency experience exercise-induced myalgias.

Second, chronic administration of adenosine can lead to desensitization of adenosine receptors (77–81). On the contrary, intermittent application of A1 receptor agonist has prevented the effect of desensitization and maintained the myocardium in the preconditioned state (82). Patients with AMPD1 deficiency presumably release from their skeletal muscles considerable amounts of adenosine each time they exercise, which could result in an intermittent manner of adenosine receptors activation in the heart. Moreover, this activation is likely to be timed during physical activity that impose increased demand on the cardiac muscle as

well, offering additional protection against ischemia. Indeed, adenosine-enhanced ischemic preconditioning (adenosine administered simultaneously with preconditioning periods of ischemia) has been shown in experiments to convey a greater degree of protection than either preconditioning stimulus alone (83, 84). As shown by the example of second window of preconditioning, the cardioprotective effect of short-term adenosine receptors stimulation can be delayed up to several days after the stimulus.

In summary, we hypothesize that during physical activity of AMPD1 deficient patient adenosine is released in great amounts from skeletal muscle and possibly from the myocardium. This metabolite can activate cardiac adenosine receptors, which proceed to multiple downstream cascade with beneficial effects for the heart. Most likely, the stimulation of the adenosine receptors in AMPD1 deficiency is not chronic and thus should not lead to their desensitization and loss of the salutary effects. The promising results of Loh's and Anderson's studies encourage further investigation in order to describe adenosine kinetics and clarify the pathophysiological mechanisms responsible for the improved cardiovascular survival in patients with AMPD1 gene mutation. If we succeed in mimicking the biochemical situation that happens in the organism of these patients, we could be able to prolong life of patients with chronic heart diseases.

References

1. **Drury AN, Szent-György A.** The physiological activity of adenine compounds with especial reference to their action upon the mammalian heart. *J Physiol (London)* 1929; 68: 213–237.
2. **Schrader J.** Adenosine. A homeostatic metabolite in cardiac energy metabolism. *Circulation* 1990; 81 (1): 389–391.
3. **Ely SW, Berne RM.** Protective effects of adenosine in myocardial ischemia. *Circulation* 1992; 85 (3): 893–904.
4. **Downey JM, Liu GS, Thornton JD.** Adenosine and the anti-infarct effects of preconditioning. *Cardiovasc Res* 1993; 27 (1): 3–8.
5. **Mullane K, Bullough D.** Harnessing an endogenous cardioprotective mechanism: cellular sources and sites of action of adenosine. *J mol Cell Cardiol* 1995; 27 (4): 1041–1054.
6. **Kitakaze M, Hori M.** It is time to ask what adenosine can do for cardioprotection. *Heart Vessels* 1998; 13 (5): 211–228.
7. **Vinten-Johansen J, Thourani VH, Ronson RS, Jordan JE, Zhao ZQ, Nakamura M, Velez D, Guyton RA.** Broad-spectrum cardioprotection with adenosine. *Ann Thorac Surg* 1999; 68 (5): 1942–1948.
8. **Sommerschild HT, Kirkeboen KA.** Adenosine and cardioprotection during ischaemia and reperfusion — an overview. (In process citation). *Acta Anaesthesiol Scand* 2000; 44 (9): 1038–1055.
9. **Loh E, Rebbeck TR, Mahoney PD, DeNofrio D, Swain JL, Holmes EW.** Common variant in AMPD1 gene predicts improved clinical outcome in patients with heart failure (see comments). *Circulation* 1999; 99 (11): 1422–1425.
10. **Anderson JL, Habashi J, Carlquist JF, Muhlestein JB, Horne BD, Bair TL, Pearson RR, Hart N.** A common variant of the AMPD1 gene predicts improved cardiovascular survival in patients with coro-

- nary artery disease. (In process citation). *J Amer Coll Cardiol* 2000; 36 (4): 1248—1252.
11. **Dubey RK, Gillespie DG, Mi Z, Jackson EK.** Cardiac fibroblasts express the cAMP-adenosine pathway. *Hypertension* 2000; 36 (3): 337—342.
12. **Dubey RK, Mi Z, Gillespie DG, Jackson EK.** Cyclic AMP-adenosine pathway inhibits vascular smooth muscle cell growth. *Hypertension* 1996; 28 (5): 765—771.
13. **Darvish A, Pomerantz RW, Zografides PG, Metting PJ.** Contribution of cytosolic and membrane-bound 5'-nucleotidases to cardiac adenosine production. *Amer J Physiol* 1996; 27 (5): H2162—H2167.
14. **Darvish A, Metting PJ.** Purification and regulation of an AMP-specific cytosolic 5'-nucleotidase from dog heart. *Amer J Physiol* 1993; 264 (5): H1528—H1534.
15. **Sala-Newby GB, Skladanowski AC, Newby AC.** The mechanism of adenosine formation in cells. Cloning of cytosolic 5'-nucleotidase-I. *J Biol Chem* 1999; 274 (25): 17789—17793.
16. **Sala-Newby GB, Freeman NV, Skladanowski AC, Newby AC.** Distinct roles for recombinant cytosolic 5'-nucleotidase-I and -II in AMP and IMP catabolism in COS-7 and H9c2 rat myoblast cell lines. *J Biol Chem* 2000; 275 (16): 11666—11671.
17. **Misumi Y, Ogata S, Hirose S, Ikehara Y.** Primary structure of rat liver 5'-nucleotidase deduced from the cDNA. Presence of the COOH-terminal hydrophobic domain for possible post-translational modification by glycopospholipid. *J Biol Chem* 1990; 265 (4): 2178—2183.
18. **Mlodzik K, Loffing J, Le Hir M, Kaissling B.** Ecto-5'-nucleotidase is expressed by pericytes and fibroblasts in the rat heart. *Histochem Cell Biol* 1995; 103 (3): 227—236.
19. **Pearson JD, Carleton JS, Gordon JL.** Metabolism of adenine nucleotides by ectoenzymes of vascular endothelial and smooth-muscle cells in culture. *Biochem J* 1980; 190 (2): 421—429.
20. **Skladanowski AC, Smolenski RT, Tavenier M, de Jong JW, Yacoub MH, Seymour AM.** Soluble forms of 5'-nucleotidase in rat and human heart. *Amer J Physiol* 1996; 270 (4): H1493—H1500.
21. **Schwartz LM, Bukowski TR, Revkin JH, Bassingthwaight JB.** Cardiac endothelial transport and metabolism of adenosine and inosine. *Amer J Physiol* 1999; 277 (3): H1241—H1251.
22. **Raatikainen MJ, Peuhkurinen KJ, Hassinen IE.** Contribution of endothelium and cardiomyocytes to hypoxia-induced adenosine release. *J Mol Cell Cardiol* 1994; 26 (8): 1069—1080.
23. **Mattig S, Deussen A.** Significance of adenosine metabolism of coronary smooth muscle cells. *Amer J Physiol Heart Circulat Physiol* 2001; 280 (1): H117—H124.
24. **Deussen A, Moser G, Schrader J.** Contribution of coronary endothelial cells to cardiac adenosine production. *Pflugers Arch* 1986; 406 (6): 608—614.
25. **Pearson JD, Gordon JL.** Vascular endothelial and smooth muscle cells in culture selectively release adenine nucleotides. *Nature* 1979; 281 (5730): 384—386.
26. **Kroll K, Decking UK, Dreikorn K, Schrader J.** Rapid turnover of the AMP-adenosine metabolic cycle in the guinea pig heart. *Circulat Res* 1993; 73 (5): 846—856.
27. **Deussen A, Stappert M, Schafer S, Kelm M.** Quantification of extracellular and intracellular adenosine production: understanding the transmembranous concentration gradient. *Circulation* 1999; 99 (15): 2041—2047.
28. **Rubin LJ, Johnson LR, Dodam JR, Dhalla AK, Magliola L, Laughlin MH, Jones AW.** Selective transport of adenosine into porcine coronary smooth muscle. *Amer J Physiol Heart Circulat Physiol* 2000; 279 (3): H1397—H1410.
29. **Deussen A.** Quantitative integration of different sites of adenosine metabolism in the heart. (In process citation). *Ann Biomed Engl* 2000; 28 (8): 877—883.
30. **Deussen A.** Metabolic flux rates adenosine in the heart. (In process citation). *Naunyn Schmiedebergs Arch Pharmacol* 2000; 362 (4—5): 351—636.
31. **Kitakaze M, Hori M, Takashima S, Sato H, Inoue M, Kamada T.** Ischemic preconditioning increases adenosine release and 5'-nucleotidase activity during myocardial ischemia and reperfusion in dogs. Implications for myocardial salvage. *Circulation* 1993; 87 (1): 208—215.
32. **Kitakaze M, Hori M, Morioka T, Minamino T, Takashima S, Okazaki Y, Node K, Komamura K, Iwakura K, Itoh T.** Alpha 1-adrenoceptor activation increases ecto-5'-nucleotidase activity and adenosine release in rat cardiomyocytes by activating protein kinase C. *Circulation* 1995; 91 (8): 2226—2234.
33. **Sparks HV, Jr, Bardenheuer H.** Regulation of adenosine formation by the heart. *Circulat Res* 1986; 58 (2): 193—201.
34. **Bardenheuer H, Schrader J.** Supply-to-demand ratio for oxygen determines formations of adenosine by the heart. *Amer J Physiol* 1986; 250 (2): H173—H180.
35. **Fenton RA, Dobson JG, Jr.** Measurement by fluorescence of interstitial adenosine levels in normoxic, hypoxic, and ischemic perfused rat hearts. *Circulat Res* 1987; 60 (2): 177—184.
36. **Goto M, Cohen MV, Van Wylen DG, Downey JM.** Attenuated purine production during subsequent ischemia in preconditioned rabbit myocardium is unrelated to the mechanism of protection. *J Mol Cell Cardiol* 1996; 28 (3): 447—454.
37. **Sollevi A.** Cardiovascular effects of adenosine in man; possible clinical implications. *Prog Neurobiol* 1986; 27 (4): 319—349.
38. **Funaya H, Kitakaze M, Node K, Minamino T, Komamura K, Hori M.** Plasma adenosine levels increase in patients with chronic heart failure. *Circulation* 1997; 95 (6): 1363—1365.
39. **Berne RM.** The role of adenosine in the regulation of coronary blood flow. *Circulat Res* 1980; 47 (6): 807—813.
40. **Newby AC.** Adenosine and the concept of „retaliatory metabolites“. *Trends Biochem Sci* 1984; 9: 42—44.
41. **Berne RM.** Cardiac nucleotides in hypoxia: Possible role in regulation of coronary blood flow. *Amer J Physiol* 1963; 204: 317—322.
42. **Murry CE, Jennings RB, Reimer KA.** Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* 1986; 74 (5): 1124—1136.
43. **Kloner RA, Jennings RB.** Consequences of brief ischemia: stunning, preconditioning, and their clinical implications: part I. *Circulation* 2001; 104 (24): 2981—2989.
44. **Woolfson RG, Patel VC, Yellon DM.** Pre-conditioning with adenosine leads to concentration-dependent infarct size reduction in the isolated rabbit heart. *Cardiovasc Res* 1996; 31 (1): 148—151.

45. **Leesar MA, Stoddard M, Ahmad M, Broadbent J, Bolli R.** Preconditioning of human myocardium with adenosine during coronary angioplasty. *Circulation* 1997; 85 (11): 2500–2507.
46. **Lasley RD, Mentzer RM, Jr.** Adenosine improves recovery of postischemic myocardial function via an adenosine A₁ receptor mechanism. *Amer J Physiol* 1992; 263 (5): H1460–H1465.
47. **Thornton CF, Liu GS, Olsson RA, Downey JM.** Intravenous pretreatment with A₁-selective adenosine analogues protects the heart against infarction. *Circulation* 1992; 85 (2): 659–665.
48. **Toombs GF, McGee S, Johnston WE, Vinten-Johansen J.** Myocardial protective effects of adenosine. Infarct size reduction with pretreatment and continued receptor stimulation during ischemia. *Circulation* 1992; 86 (3): 986–994.
49. **Morrison RR, Jones R, Byford AM, Stell AR, Peart J, Headrick JP, Matherne GP.** Transgenic overexpression of cardiac A₁ adenosine receptors mimics ischemic preconditioning. *Amer J Physiol Heart Circulat Physiol* 2000; 279 (3): H1017–H1078.
50. **Liang BT, Jacobson KA.** A physiological role of the adenosine A₃ receptor: sustained cardioprotection. *Proc Natl Acad Sci USA* 1998; 95 (12): 6995–6999.
51. **Thourani VH, Nakamura M, Ronson RS, Jordan JE, Zhao ZQ, Levy JH, Szlam F, Guyton RA, Vinten-Johansen J.** Adenosine A₃-receptor stimulation attenuates postischemic dysfunction through K(ATP) channels. *Amer J Physiol* 1999; 277 (1): H228–H235.
52. **Thourani VH, Ronson RS, Jordan JE, Guyton RA, Vinten-Johansen J.** Adenosine A₃ pretreatment before cardioplegic arrest attenuates postischemic cardiac dysfunction. *Ann Thorac Surg* 1999; 67 (6): 1732–1737.
53. **Harrison GJ, Gerniway RJ, Peart J, Berr SS, Ashton K, Regan S, Paul MG, Headrick JP.** Effects of A₃ adenosine receptor activation and gene knock-out in ischemic-reperfused mouse heart. *Cardiovasc Res* 2002; 53 (1): 147–155.
54. **Peart J, Flood A, Linden J, Matherne GP, Headrick JP.** Adenosine-mediated cardioprotection in ischemic-reperfused mouse heart. *J Cardiovasc Pharmacol* 2002; 39 (1): 117–129.
55. **Baxter GF, Marber MS, Patel VC, Yellon DM.** Adenosine receptor involvement in a delayed phase of myocardial protection 24 hours after ischemic preconditioning. *Circulation* 1994; 90 (6): 2993–3000.
56. **Dana A, Skarli M, Papakrivopoulou J, Yellon DM.** Adenosine A₁ receptor induced delayed preconditioning in rabbits: induction of p38 mitogen-activated protein kinase activation and Hsp27 phosphorylation via a tyrosine kinase- and protein kinase C-dependent mechanism. *Circulat Res* 2000; 86 (9): 989–997.
57. **Carroll R, Yellon DM.** Delayed cardioprotection in a human cardiomyocyte-derived cell line: the role of adenosine, p38MAP kinase and mitochondrial KATP. (In process citation). *Basic Res Cardiol* 2000; 95 (3): 243–249.
58. **Ikeda U, Kurosaki K, Shimpo M, Okada K, Saito T, Shimada K.** Adenosine stimulates nitric oxide synthesis in rat cardiac myocytes. *Amer J Physiol* 1997; 273 (1): H59–H65.
59. **Dana A, Jonassen AK, Yamashita N, Yellon DM.** Adenosine A₁ receptor activation induces delayed preconditioning in rats mediated by manganese superoxide dismutase. *Circulation* 2000; 101 (24): 2841–2848.
60. **Liu GS, Thornton J, Van Winkle DM, Stanley AW, Olsson RA, Downey JM.** Protection against infarction afforded by preconditioning is mediated by A₁ adenosine receptors in rabbit heart. *Circulation* 1991; 84 (1): 350–356.
61. **Cleveland JC, Jr, Meldrum DR, Rowland RT, Banerjee A, Harken AH.** Adenosine preconditioning of human myocardium is dependent upon the ATP-sensitive K⁺ channel. *J Mol Cell Cardiol* 1997; 29 (1): 175–182.
62. **Pain T, Yang XM, Critz SD, Yue Y, Nakano A, Liu GS, Heusch G, Cohen MV, Downey JM.** Opening of mitochondrial K(ATP) channels triggers the preconditioned state by generating free radicals. *Circulat Res* 2000; 87 (6): 460–466.
63. **Miura T, Liu Y, Kita H, Ogawa T, Shimamoto K.** Roles of mitochondrial ATP-sensitive K channels and PKC in anti-infarct tolerance afforded by adenosine A₁ receptor activation. *J Amer Coll Cardiol* 2000; 35 (1): 238–245.
64. **Tanno M, Tsuchida A, Nozawa Y, Matsumoto T, Hasegawa T, Miura T, Shimamoto K.** Roles of tyrosine kinase and protein kinase C in infarct size limitation by repetitive ischemic preconditioning in the rat. *J Cardiovasc Pharmacol* 2000; 35 (3): 345–352.
65. **Weinbrenner C, Liu GS, Cohen MV, Downey JM.** Phosphorylation of tyrosine 182 of p38 mitogen-activated protein kinase correlates with the protection of preconditioning in the rabbit heart. *J Mol Cell Cardiol* 1997; 29 (9): 2383–2391.
66. **Takano H, Manchikalapudi S, Tang XL, Qiu Y, Rizvi A, Jadoon AK, Zhang Q, Bolli R.** Nitric oxide synthase is the mediator of late preconditioning against myocardial infarction in conscious rabbits. *Circulation* 1998; 98 (5): 441–449.
67. **Marber MS, Matchman DS, Walker JM, Yellon DM.** Cardiac stress protein elevation 24 hours after brief ischemia or heat stress is associated with resistance to myocardial infarction. *Circulation* 1993; 88 (3): 1264–1272.
68. **Sakamoto K, Urushidani T, Nagao T.** Translocation of HSP27 to sarcomere induced by ischemic preconditioning is isolated rat hearts. *Biochem Biophys Res Commun* 2000; 269 (1): 137–142.
69. **Mahaffey KW, Puma JA, Barbagelata NA, DiCarli MF, Leesar MA, Browne KF, Eisenberg PR, Bolli R, Casas AC, Molina-Viamonte V, Orlandi C, Blevins R, Gibbons RJ, Califf RM, Granger CB.** Adenosine as an adjunct to thrombolytic therapy for acute myocardial infarction: results of a multicenter, randomized, placebo-controlled trial: the acute myocardial infarction study of adenosine (AMISTAD) trial. *J Amer Coll Cardiol* 1999; 34 (6): 1711–1720.
70. **Heidland UE, Heintzen MP, Michel CJ, Strauer BE.** Effect of adjunctive intracoronary adenosine on myocardial ischemia, hemodynamic function and left ventricular performance during percutaneous transluminal coronary angioplasty: clinical access to ischemic preconditioning? (In process citation). *Coron Artery Dis* 2000; 11 (5): 421–428.
71. **Teoh LK, Grant R, Hulf JA, Pugsley WB, Yellon DM.** The effect of preconditioning (ischemic and pharmacological) on myocardial necrosis following coronary artery bypass graft surgery. *Cardiovasc Res* 2002; 53 (1): 175–180.
72. **Mentzer RM, Jr, Rahko PS, Molina-Viamonte V, Canver CC, Chopra PS, Love RB, Cook TD, Hegge JO, Lasley RD.** Safety, tolerance, and efficacy of adenosine as an additive to blood cardioplegia in humans during coronary artery bypass surgery. *Amer J Cardiol* 1997; 79 (12A): 38–43.
73. **Chauhan S, Wasir HS, Bhan A, Rao BH, Saxena N, Venugopal P.** Adenosine for cardioplegic induction: a comparison with St Thomas solution. *J Cardiothorac Vasc Anesth* 2000; 14 (1): 21–24.

- 74. Tanfani F, Kulawiak D, Kossowska E, Preis JP, Zydowo MM, Sarkissova E, Bertoli E, Wozniak M.** Structural-functional relationships in pig heart AMP-deaminase in the presence of ATP, orthophosphate, and phosphatidate bilayers. *Mol Genet Metab* 1998; 65 (1): 51—58.
- 75. Feldman AM, Wagner DR, McNamara DM.** AMPD1 gene mutation in congestive heart failure: new insights into the pathobiology of disease progression (editorial). *Circulation* 1999; 99 (11): 1397—1399.
- 76. Sabina RL, Swain JL, Olanow CW, Bradley WG, Fishbein WN, DiMauro S, Holmes EW.** Myoadenylate deaminase deficiency. Functional and metabolic abnormalities associated with disruption of the purine nucleotide cycle 1. *J Clin Invest* 1984; 73 (3): 720—730.
- 77. Longabaugh JP, Didsbury J, Spiegel A, Stiles GL.** Modification of the rat adipocyte A1 adenosine receptor-adenylate cyclase system during chronic exposure to an A1 adenosine receptor agonist: alterations in the quantity of GS alpha and Gi alpha are not associated with changes in their mRNAs. *Mol Pharmacol* 1989; 36 (5): 681—688.
- 78. Liang BT, Donovan LA.** Differential desensitization of A1 adenosine receptor-mediated inhibition of cardiac myocyte contractility and adenylate cyclase activity. Relation to the regulation of receptor affinity and density. *Circulat Res* 1990; 67 (2): 406—414.
- 79. Lee HT, Thompson CI, Hernandez A, Lewy JM, Belloni FL.** Cardiac desensitization to adenosine analogues after prolonged R-PIA infusion in vivo. *Amer J Physiol* 1993; 265 (6): H1916—H1927.
- 80. Tsuchida A, Thompson R, Olsson RA, Downey JM.** The anti-infarct effect of an adenosine A1-selective agonist is diminished after prolonged infusion as is the cardioprotective effect of ischaemic preconditioning in rabbit heart. *J Mol Cell Cardiol* 1994; 26 (3): 303—311.
- 81. Palmer TM, Benovic JL, Stiles GL.** Agonist-dependent phosphorylation and desensitization of the rat A3 adenosine receptor. Evidence for a G-protein-coupled receptor kinase-mediated mechanism. *J Biol Chem* 1995; 270 (49): 29607—29613.
- 82. Dana A, Baxter GF, Walker JM, Yellon DM.** Prolonging the delayed phase of myocardial protection: repetitive adenosine A1 receptor activation maintains rabbit myocardium in a preconditioned state. *J Amer Coll Cardiol* 1998; 31 (5): 1142—1149.
- 83. McCully JD, Levitsky S.** Alternatives for myocardial protection: adenosine-enhanced ischemic preconditioning. *Ann NY Acad Sci* 1999; 874: 295—305.
- 84. Stadler B, Phillips J, Toyoda Y, Federman M, Levitsky S, McCully JD.** Adenosine-enhanced ischemic preconditioning modulates necrosis and apoptosis: effects of stunning and ischemia-reperfusion. *Ann Thorac Surg* 2001; 72: 555—564.

Received November 25, 2001.

Accepted May 20, 2002.